Portland State University

PDXScholar

Environmental Science and Management Faculty Publications and Presentations

Environmental Science and Management

8-24-2021

Balanced Polymorphism Fuels Rapid Selection in an Invasive Crab Despite High Gene Flow and Low Genetic Diversity.

C K. Tepolt Woods Hole Oceanographic Institution

E D. Grosholz DepUniversity of California, Davis

Catherine E. de Rivera Portland State University

G M. Ruiz Smithsonian Environmental Research Center, Smithsonian Institution

Follow this and additional works at: https://pdxscholar.library.pdx.edu/esm_fac

Part of the Environmental Sciences Commons Let us know how access to this document benefits you.

Citation Details

Tepolt, C. K., Grosholz, E. D., de Rivera, C. E., & Ruiz, G. M. (2021). Balanced polymorphism fuels rapid selection in an invasive crab despite high gene flow and low genetic diversity. *Molecular Ecology*, mec.16143. https://doi.org/10.1111/mec.16143

This Post-Print is brought to you for free and open access. It has been accepted for inclusion in Environmental Science and Management Faculty Publications and Presentations by an authorized administrator of PDXScholar. Please contact us if we can make this document more accessible: pdxscholar@pdx.edu.



	1								
	2	DR. CAROLYN TEPOLT (Orcid ID : 0000-0002-7062-3452)							
	3								
(5	Article type : Original Article							
	6								
	7	Delevered vehicles field would be be the in an investigation and							
	8	Balanced polymorphism fuels rapid selection in an invasive crab							
	9	despite high gene flow and low genetic diversity							
	10								
	11	CK Tepolt ^{1,*} , ED Grosholz ² , CE de Rivera ³ , GM Ruiz ⁴							
	12								
	13	¹ Department of Biology, Woods Hole Oceanographic Institution, 266 Woods Hole Road,							
	14	Woods Hole, MA 02543							
	15	² Department of Environmental Science and Policy, University of California, Davis, CA 95616							
	16	³ Department of Environmental Science and Management, Portland State University, Box 751,							
	17	Portland, OR 97207							
	18	⁴ Smithsonian Environmental Research Center, Smithsonian Institution, 647 Contees Wharf							
	19	Road, Edgewater, MD 21037							
	20								
	21	*Corresponding Author:							
	22	Carolyn Tepolt							
	23	Department of Biology							
	24	Woods Hole Oceanographic Institution							
	25	266 Woods Hole Road, MS #33							
	26	508-289-3357							
	27	ctepolt@whoi.edu							
	28								
	29								
	30	Keywords: rapid adaptation, invasive species, island of divergence, seascape genomics,							
		This article has been accepted for publication and undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the <u>Version of Record</u> . Please cite this article as <u>doi: 10.1111/MEC.16143</u>							

1 balanced polymorphism

32 Abstract:

33

34 Adaptation across environmental gradients has been demonstrated in numerous systems with extensive dispersal, despite high gene flow and consequently low genetic structure. The 35 speed and mechanisms by which such adaptation occurs remain poorly resolved, but are 36 37 critical to understanding species spread and persistence in a changing world. Here, we investigate these mechanisms in the European green crab *Carcinus maenas*, a globally 38 distributed invader. We focus on a northwestern Pacific population that spread across >12 39 40 degrees of latitude in 10 years from a single source, following its introduction <35 years ago. Using six locations spanning >1,500 km, we examine genetic structure using 9,376 Single 41 Nucleotide Polymorphisms (SNPs). We find high connectivity among five locations, with 42 significant structure between these locations and an enclosed lagoon with limited connectivity 43 to the coast. Among the five highly connected locations, the only structure observed was a 44 45 cline driven by a handful of SNPs strongly associated with latitude and winter temperature. These SNPs are almost exclusively found in a large cluster of genes in strong linkage 46 disequilibrium that was previously identified as a candidate for cold tolerance adaptation in 47 48 this species. This region may represent a balanced polymorphism that evolved to promote 49 rapid adaptation in variable environments despite high gene flow, and which now contributes to successful invasion and spread in a novel environment. This research suggests an answer 50 51 to the paradox of genetically depauperate yet successful invaders: populations may be able to adapt via a few variants of large effect despite low overall diversity. 52

53 Introduction

54

55 In the ocean, where many species are characterized by large population sizes, long-distance planktonic dispersal, and broad ranges (Kinlan & Gaines, 2003; Palumbi & Pinsky, 2014), 56 there has been a classical assumption of genetic homogeneity and little persistent 57 differentiation (Hedgecock, 1986). Recently, however, both population genomics and 58 comparative physiology have uncovered evidence of genetic selection and functional 59 differences among widespread populations living across varied marine environments (Sanford 60 61 & Kelly, 2011; Pespeni & Palumbi, 2013). Likewise, genetic studies and associated modeling have detected subtle genetic structure driven by oceanography in species that disperse 62 widely (Galindo, Olson, & Palumbi, 2006; White et al., 2010; Xuereb et al., 2018). This 63 increasing evidence for differentiation in the sea begs the question of how quickly, and 64 through which mechanisms, marine species may cope with rapidly changing environmental 65 conditions (Munday, Warner, Monro, Pandolfi, & Marshall, 2013). Introduced species, which 66 in many cases establish and thrive in novel habitats, offer the opportunity to examine these 67 questions in the context of the natural environment (Blackburn, 2008; Lee, Kiergaard, 68 69 Gelembiuk, Eads, & Posavi, 2011).

70

71 Marine species exhibit a spectrum of evolutionary mechanisms based in part on their 72 dispersal. Local adaptation in the classical sense is restricted to species with relatively limited 73 dispersal, which facilitates the selection and retention of adaptive alleles within a population 74 (Kawecki & Ebert, 2004). Relatively isolated populations are also likely to diverge due to neutral processes, as genetic drift changes allele frequencies across the genome (Ellingson & 75 Krug, 2016; Prunier, Dubut, Chikhi, & Blanchet, 2017). On the other end of this evolutionary 76 77 spectrum lie open marine systems, where alleles are continually exported from each location 78 to a mixed pool of larvae that may settle in environments far different from their sources. In 79 this dynamic, balanced polymorphism is favored, and adaptive variation is maintained within 80 the population as a whole (Sanford & Kelly, 2011). These adaptive alleles mix as larvae disperse, and the environmental conditions they encounter as they recruit can result in strong 81 and rapid selection that culls less-fit alleles from the local population (Sotka, 2012). This 82 phenomenon has been described largely in the context of maintaining differentiation across 83

small-scale environmental differences year after year in systems where the scale of dispersal 84 far exceeds the scale of selection. For example, strong selection to salinity appears to have 85 maintained an enzymatic cline in mussels along Long Island Sound (Koehn, Newell, & 86 Immermann, 1980), and microhabitat differences across tidal heights maintain balanced 87 polymorphism in limpet populations (Schmidt, Bertness, & Rand, 2000). Such examples may 88 better reflect the realized capacity of highly dispersive marine species to adapt to stressors 89 across complex oceanographic regimes than studies of strict local adaptation (Véliz, 90 Duchesne, Bourget, & Bernatchez, 2006). 91

As species expand into new environments, the process of adaptation may be mediated by the 93 complex demographic effects that often occur at range edges (Bridle & Vines, 2007; Chuang 94 & Peterson, 2016). Expanding populations are frequently characterized by sequential 95 bottlenecks and losses of genetic diversity caused by small groups of colonizing organisms 96 (Eckert, Samis, & Lougheed, 2008; White, Perkins, Heckel, & Searle, 2013; Bors, Herrera, 97 Morris, & Shank, 2019). The success of some such populations, which multiply and spread 98 despite low genetic diversity, has been coined the "genetic paradox of invasions" (Roman & 99 100 Darling, 2007). These bottlenecks can also lead to increased stochasticity at range edges, causing allele surfing and other distinctive genetic patterns (Excoffier & Ray, 2008). Gene 101 102 flow plays a substantial role in this process. In some cases, low-diversity edge populations may be evolutionarily limited by a lack of gene flow (Sexton, Strauss, & Rice, 2011; 103 104 Takahashi et al., 2016), while in others, semi-isolation at range edges permits rapid evolution in organisms at the expansion front (Phillips, Brown, Webb, & Shine, 2006; Kilkenny & 105 Galloway, 2012; Szücs et al., 2017). High dispersal and large populations may also facilitate 106 107 species persistence and expansion, if functional diversity can be maintained and guickly spread throughout the expanding range (Rius & Darling, 2014). The maintenance of such 108 diversity can in theory provide the raw substrate for adaptation and permit extremely quick 109 evolutionary response to shifting conditions (Tigano & Friesen, 2016; Llaurens, Whibley, & 110 111 Joron, 2017). However, to date, the relative contributions of selection and drift as populations establish in novel environments have not been explored empirically in high gene flow 112 systems. 113

114

92

The European green crab (*Carcinus maenas*) along the northeast Pacific coastline presents 115 an ideal test case for untangling the dynamics of rapid marine adaptation and differentiation 116 with high potential gene flow. In this region, the species established an initial population in 117 San Francisco Bay by 1990 (Carlton & Cohen, 2003), and spread >1,500 km to Vancouver 118 Island in <10 years despite deriving from a single, significantly bottlenecked source (Tepolt et 119 al., 2009). Green crabs advanced along the coast and primarily to the north, reaching 120 northern California by 1995, southern Oregon by 1997, and Vancouver Island, British 121 Columbia by 1998 (Behrens Yamada & Gillespie, 2008; Fig. 1). This rapid expansion was 122 associated with extremely strong positive El Niño-Southern Oscillation (ENSO) indices in 123 1997-1998 that promoted high reproductive output, northward transport, and coastal retention 124 of larvae (Behrens Yamada et al., 2005; Behrens Yamada & Kosro, 2010; See & Feist, 2010). 125 Importantly, in the northeast Pacific, C. maenas are found almost exclusively in shallow 126 127 waters of protected embayments and not along the exposed outer coast between bays, resulting in a disjunct distribution of green crab populations. This habitat distribution is similar 128 to its introduced range in South Africa, where the rocky coast is also subject to high-energy 129 wave action (Hampton & Griffiths, 2007), but differs from its more continuous distribution in 130 131 other global regions (Carlton & Cohen, 2003). The species also has a relatively wide 132 environmental tolerance and diet breadth, along with a 30-75 day pelagic larval duration (Dawirs, 1985), which have contributed to its spread and establishment in six introduced 133 134 regions across five continents (Carlton & Cohen, 2003; Hidalgo, Barón, & Orensanz, 2005). 135 The importance of ENSO events in the spread and abundance of northeast Pacific C. maenas 136

suggests that both temperature and local oceanography may play substantial roles in 137 structuring the population. Decades of field surveys of abundance in both the northwest 138 Atlantic and northeast Pacific support the importance of temperature during early 139 development in driving recruitment strength: cold winters have been associated with weaker 140 recruitment and smaller cohorts of crabs than milder years (Behrens Yamada & Kosro, 2010; 141 Welch, 1968). A global physiological study demonstrated population-level differences in adult 142 heat and cold tolerance consistent with local adaptation (Tepolt & Somero, 2014). 143 Subsequently, transcriptomic work has identified genetic markers associated with 144 temperature tolerance on a population level (Tepolt & Palumbi, 2020). Like many marine 145

species, *C. maenas* larvae have shown narrower temperature tolerances than adults in
laboratory trials, suggesting that thermal tolerance at early life stages may be particularly
important in shaping crab populations across different years (Dawirs, 1985; de Rivera et al.,
2007).

150

Here, we use transcriptome-derived SNPs from C. maenas populations in the northeast 151 Pacific to test the roles of connectivity and selection in shaping the population structure of this 152 highly dispersive and recently introduced species. Using six sites spanning over 1,500 km of 153 coastline, we examine population structure and relative migration to elucidate connectivity 154 among embayments across the species' northeast Pacific range. For a few sites, we have 155 temporal samples spanning 2-5 years, which we use to examine the stability of population 156 structure over time. Finally, we test for candidate genes for selection across a thermal 157 latitudinal gradient, comparing these candidates to genes identified in a prior global study of 158 the genetic basis of thermal tolerance differences in the species. As this population was 159 founded <35 years ago from a single source, our data represent patterns of divergence and 160 selection that have arisen in under 20 generations. 161

162

163 Materials and Methods

164

165 Sample Collection

Twelve crabs were sampled from each of six sites along the northeast Pacific range of *C. maenas* in 2015-2016 (Figure 1). Two of these sites (Seadrift Lagoon and San Francisco Bay,
CA, USA) were sampled in both years, while the remaining sites were sampled once. We also
reanalyzed raw sequence data from a prior study of crabs collected in 2011 from two sites
(Seadrift Lagoon, CA, USA and Barkley Sound, BC, Canada; Tepolt & Palumbi 2015). Crabs
were collected by hand or trap, and hearts were dissected and stored in RNALater at -80°C.

173 Extraction & Sequencing

Total RNA was extracted from cardiac tissue using TRIzol (Invitrogen, Carlsbad, CA, USA)
with 1-bromo-3-chloropropane (Simms, Cizdziel, & Chomczynski, 1993). RNA was quantified
using the broad-range RNA assay on a Qubit 3.0 fluorometer (Invitrogen), and up to 4 µg of

- 177 RNA was used to prepare individually-barcoded cDNA libraries with Ilumina's TruSeq
- 178 Stranded mRNA Library Prep Kit (Illumina, San Diego, CA, USA). Libraries were sent to the
- 179 University of California Berkeley's Genomics Sequencing Laboratory, where they were
- 180 quantified and pooled into groups of 16 multiplexed samples run on five lanes of an Illumina
- 181 HiSeq 4000 in 50-bp single-end reads.
- 182
- 183 Sequence Processing and SNP Identification
- 184 Raw sequences were cleaned and trimmed using Trim Galore! v0.6.4
 185 (http://www.bioinformatics.babraham.ac.uk/projects/trim_galore/), a wrapper for Cutadapt v2.6
- (Martin 2011). A nucleotide call quality cutoff of Phred \geq 20 was used, and reads \leq 20bp after 186 adapter removal and quality trimming were discarded. A published C. maenas cardiac 187 transcriptome was used as a reference (Tepolt & Palumbi 2015), after an expression-based 188 189 screening to remove poorly-supported contigs and reduce computational load. Briefly, we mapped trimmed and clipped reads from Tepolt & Palumbi 2015 back to the reference 190 transcriptome using salmon v1.2.1 (Patro, Duggal, Love, Irizarry, & Kingsford, 2017). We 191 retained only contigs with TPM >1, and re-annotated these contigs using EnTAP v 0.9.1 (Hart 192 193 et al. 2020), comparing all 6 reading frames against the Swissprot, TrEMBL, and nr protein databases (downloaded March 2020). Annotations to Decapoda were prioritized with the 194 195 program's '--taxon' flag. Contigs with a clear taxonomic mismatch to decapods (e.g., bacteria, 196 green plants, fungi, etc.), as well as all likely mitochondrial and ribosomal contigs, were 197 identified and removed from the project after alignment to minimize non-target mapping. This 198 resulted in a clean reference transcriptome of 25,552 nuclear contigs.
- 199

Cleaned reads were mapped back to the *C. maenas* cardiac transcriptome using Bowtie2
v2.4.1 with default settings (Langmead & Salzberg 2013). Picard v2.22.0 was used to sort
reads, identify and mark duplicate read sequences, and index the resulting bam files
(http://broadinstitute.github.io/picard). The Genome Analysis Toolkit (GATK) v4.1.7.0 was
used to identify and genotype biallelic SNPs (McKenna et al. 2010; DePristo et al. 2011).

Across all 120 samples (24 from 2011 and 96 from 2015-2016), GATK identified 163,261
 biallelic SNPs with Phred quality scores ≥20. We identified high-quality, well-supported SNPs

for downstream analyses using a custom python script that retained only individual genotypes 208 with Phred \geq 20 and supported by \geq 5 reads. We excluded SNPs missing high-quality 209 210 genotypes at ≥ 4 individuals for any site-by-year samples of 12 individuals. SNPs with heterozygosity ≥ 0.7 (to screen out obvious paralogs), and SNPs where the alternate allele 211 was observed only once across all individuals (to minimize bias by potential sequencing 212 errors). In total, 9,376 SNPs had high-quality genotypes for ≥ 8 individuals per group and were 213 214 retained for downstream processing. All 120 individuals had high-guality genotypes at >80% 215 of these SNPs.

216

217 Identification of Putative Inversion Polymorphisms

We explored the relationship of the 9,376 high-quality SNPs to identify potential inversion
polymorphisms and other disproportionately large groups of SNPs in linkage disequilibirum
(LD). Pairwise R² was calculated across all SNPs using the --geno-r2 and --interchrom-genor2 options in vcftools v0.1.16 (Danecek et al. 2011). We then used the R package LDna v0.64
to identify networks of SNPs in large, compact clusters, setting minimum edges to 45
(expected for 10 closely-linked SNPs) and phi to 15 (Kemppainen et al. 2015).

225 We identified one large outlier LD cluster, which contained 168 SNPs from 56 different contigs. To further investigate this cluster, we explored the relationship between member 226 SNPs using Principal Components Analysis (PCA) in smartPCA, implemented in Eigensoft 227 v7.2.1 (Price et al., 2006). We used the 116 individuals that had <20% missing genotypes 228 across the 168 SNPs in this cluster. This PCA separated individuals into three clear groups 229 along PC1, and we calculated F_{IS} within each of these groups to determine relative 230 heterozygosity (Kemppainen et al. 2015). These analyses strongly suggested an inversion 231 polymorphism (see Results below), so for clarity we refer to this group of 168 SNPs as an 232 "inferred inversion" throughout the rest of the manuscript. To compare the impact this putative 233 inversion had on overall population structure, we also ran a PCA on the same 116 individuals 234 using the full SNP set both with and without the 168 SNPs in the inferred inversion (N = 9.376) 235 and 9,208 SNPs, respectively). 236

237

238 Genetic Structure and Diversity

SNPs were separately screened to identify all sets of markers in linkage disequilibrium (LD) 239 and generate a set of independent SNPs for population genomics. Pairwise R² values 240 (calculated above) were used to identify groups of two or more SNPs in LD at $R^2 \ge 0.8$ using 241 the R package igraph v1.2.4.2 (Csardi & Nepusz 2006), and then all but one SNP in each 242 group was removed, leaving a set of 6,848 independent SNPs. The one SNP retained from 243 each group had the highest number of high-quality genotypes, with lower-coverage SNPs 244 within an LD group preferentially removed. We use the term "independent" to indicate that 245 these SNPs have been screened to remove those in strong LD, but note that these SNPs 246 may be in LD at lower levels so are not all truly independent. Similarly, this set of 6,848 247 independent SNPs retained 54 of the 168 SNPs in the inferred inversion which were in lower 248 levels of LD with each other (R²: 0.29-0.79). 249

250

251 Basic descriptive statistics were calculated for each site-by-year sample using the set of 6,848 independent SNPs (Table 1). Allelic richness (Ar) and private allelic richness (pAr) were 252 determined using ADZE v1.0 (Szpiech, Jakobsson, & Rosenberg, 2008). The R package 253 genepop v1.1.7 was used to calculate observed and expected heterozygosity (Ho and He), 254 255 and to calculate the inbreeding coefficient (F_{IS}) and test for heterozygote excess or deficiency 256 using Hardy-Weinberg tests (Rousset 2008). The number of polymorphic SNPs in each sample was determined using Arlequin v.3.5.2.2 (linux core implementation; Excoffier & 257 258 Lischer 2010).

259

To identify a subset of putatively neutral SNPs with which to examine population structure 260 unconfounded by selection, we used BayPass v2.2 in the core model (Gautier et al. 2015), 261 with each site-by-year sample treated as its own group. We assessed potential outliers using 262 a simulated pseudo-observed data set of 6,848 SNPs with the parameters of the real data to 263 set a 10% false discovery rate (FDR) threshold for SNP XtX. This conservative threshold was 264 chosen to avoid retaining SNPs under weak selection, thus yielding a set of SNPs more likely 265 to be truly neutral. This frequency-based approach removed all but one of the SNPs later 266 identified as a candidate for environmental association (see below). 267

268

Genetic structure for both the independent (N = 6,848) and putatively neutral (N = 6,311) SNP

sets was assessed with smartPCA. Pairwise F_{ST} was calculated between all site-by-year
groups according to Weir & Cockerham's (1984) approach using the R package 'StAMPP'
v1.6.1 with both the independent and putatively neutral SNP sets (Pembleton, Cogan, &
Forster, 2013). Significance was assessed using 10,000 permutations, and resulting p-values
were adjusted for multiple tests using a Benjamini-Hochberg false discovery rate correction
(Benjamini & Hochberg 1995).

276

We included only the most recent temporal sample from each site for an analysis of relative 277 278 migration using the putatively neutral SNP set (Table 1). Symmetry and relative magnitude of migration between sites was assessed using the 'divMigrate' function in the R package 279 'diveRsity' v1.9.90, with the Nm method and 1,000 bootstraps (Sundqvist, Keenan, 280 Zackrisson, Prodöhl, & Kleinhans, 2016). This approach, which calculates relative directional 281 282 migration, was chosen because it is more robust if populations do not perfectly satisfy some of the assumptions underlying approaches to quantify an effective migration rate (e.g., island 283 model, mutation-drift equilibrium). 284

285

286 All analyses showed strong separation of a single site, Seadrift Lagoon, from all other sites 287 (see Results below). Because of this genetic distinctiveness, isolation-by-distance (IBD) analysis excluded pairwise comparisons with Seadrift Lagoon, focusing only on the remaining 288 289 five "open" sites. Pairwise F_{ST} values were used to plot IBD between sites, using along-shore distance calculated at 50km resolution with the USA map from GADM supplemented with 290 Google Maps for distances <50km (gadm.org). When comparisons spanned San Francisco 291 Bay and the Strait of Juan de Fuca, distances were calculated across the mouths of these 292 293 features. IBD was plotted using five different SNP sets, to explore different potential drivers of latitudinal structure along the coast: 1) 6,848 independent SNPs, 2) 6,311 putatively neutral 294 SNPs, 3) 54 "independent" SNPs in the inferred inversion, 4) 6,794 independent SNPs 295 excluding the 54 independent SNPs in the inferred inversion 5) 144 outlier SNPs identified 296 among the five "open" sites with BayPass (see Results). The significance of these 297 relationships was assessed using linear regression in R. 298

299

300 Markers under Selection

We identified SNPs potentially under environmental selection using BayPass and 301 Redundancy Analysis (RDA). We ran these tests using only the five open sites, excluding 302 Seadrift Lagoon, to identify markers potentially driving the observed signal of IBD and 303 latitudinal structuring along the coast. For this testing we used a single sample, collected in 304 2015-2016, from each site: 6,662 SNPs of the full 6,848 SNP set were polymorphic in these 305 five samples and were used for tests of selection. Tested covariates included site latitude (as 306 Cartesian Y-values) and winter and summer sea surface temperature (SST). Temperature 307 data were derived from NOAA's OI SST V2 High Resolution Dataset provided by the 308 309 NOAA/OAR/ESRL PSD, Boulder, Colorado, USA, from their website at https://www.esrl.noaa.gov/psd/ (Reynolds et al. 2007). Daily temperatures were averaged 310 over the 2 years prior to sampling, and were determined for the nearest 0.25° grid location to 311 each study site for January (winter) and July (summer). 312

313

In BayPass, we first ran a core model analysis to identify frequency outlier SNPs, then used the auxiliary covariate model to test for associations between allele frequencies and latitude and winter and summer SST (Gautier 2015). Covariates were scaled, and we used an Ising prior of 0 since the physical order of contigs is unknown. We considered any SNP with Bayes factor (BF) \geq 15 dB to be a candidate for selection with respect to a given covariate.

320 We ran an RDA on individual genotypes using the R package 'vegan' v2.5-6 (Oksanen et al. 2019), with missing genotypes imputed to be the most common observed. Few individual 321 genotypes were missing in our data set: genotyping was $\geq 90\%$ complete for 97.1% of SNPs, 322 and no single sampling site was missing >4 individual genotypes at any SNP. We used 323 latitude and summer temperature as covariates (RDA formula = individual genotypes ~ Y + 324 July SST); winter temperature was excluded as it was strongly correlated with latitude (R² = 325 0.79; p = 0.03). We considered any SNP >3 SD outside the mean loading for its RDA axis to 326 be a candidate for selection (e.g., Forester et al., 2018). 327

328

SNPs were considered strong candidates for selection if they were identified by both BayPass
 and RDA. Latitude is largely correlated with SST in this data set, due both to the strongly
 linear north-south arrangement of sites along the coast and to the relative coarseness of the

332 satellite-derived temperature data. Because of this confounding, we treated all SNPs

identified as candidates for association with either latitude or temperature interchangeably. To

visualize these relationships, candidate SNPs were tested for linear correlation between

335 minor allele frequency and latitude or temperature.

336

337 Potential Function of Candidate SNPs

Candidate SNPs were examined for their potential impact on protein structure, to provide an 338 initial idea of SNPs that were particularly likely to affect organismal function. Open Reading 339 Frames (ORFs) and corresponding coding sequences for all contigs were predicted using 340 OrfPredictor v3.0 (Min, Butler, Storms, & Tsang, 2005); as many contigs could not be 341 annotated, sequence data alone was used to predict ORFs. Predicted sequences were then 342 used to class the impact of a given SNP on the resulting protein sequence (untranslated, 343 344 synonymous, or non-synonymous) using a custom python script. Of the 9,376 high-quality SNPs, we predicted that 4,339 were in the untranslated regions of the mRNA and two were in 345 contigs for which an ORF could not be predicted. Of the putatively coding SNPs, 3,843 were 346 predicted to be synonymous and 1,192 to be non-synonymous. We examined SNP 347 348 substitution patterns for all 9,376 candidate SNPs prior to LD screening. While putatively 349 linked SNPs were removed from the data set prior to selection analysis, they could potentially be driving any relationships we detected in SNPs with which they are in strong LD. 350

351

Data manipulation and plotting were done using the data.table and ggplot2 packages in R
(Wickham 2009; R Core Team 2016; Dowle & Srinivasan 2017).

354

355 **Results**

356

357 Inferred Inversion Polymorphism

Linkage disequilibrium network analysis identified one single outlier LD cluster nested in one compound outlier LD cluster (Figure 2A-B). The compound outlier LD cluster, which we refer to as an "inferred inversion", contained 168 SNPs at an LD of $R^2 \ge 0.29$. PCA of these 168 SNPs split individuals into three discrete groups along the first principal component, explaining 73.08% of variance (p = 0.001; Figure 2C). While individuals from most sites

appeared in all three groups, the left-most group contained predominantly British Columbia 363 and Seadrift Lagoon while the right-most group contained predominantly Elkhorn Slough and 364 San Francisco Bay. This pattern of three distinct groups explaining the majority of structure, 365 with no discrete partitioning of sites, is diagnostic of a region of the genome where 366 recombination is reduced (Kemppainen et al. 2015). In that case, each group represents a 367 karyotype: homozygotes on the left and right sides of PC1, and heterozygotes midway 368 between. Analysis of F_{IS} within each of these groups supports this conclusion, with values 369 near zero in the putative homozygotes (indicating Hardy-Weinberg equilibrium) and strongly 370 371 negative in the putative heterozygotes (indicating an excess of heterozygous individuals; Figure 2D). Overall F_{1S} is lower in the middle than in the left or right groups (p < 0.0001 for 372 both comparisons), while the left and right groups are not significantly different from each 373 other (p = 0.1). 374

375

Genetic structure over all SNPs showed a significant divide between Seadrift Lagoon and the other five sites, both with and without the 168 SNPs in the inferred inversion (p < 0.0001; Figure 2E-F). With the full set of 9,376 SNPs, individuals in both Seadrift Lagoon and the remaining five "open" sites were further subdivided into three clusters according to their inferred inversion genotype (p < 0.0001; Figure 2C,E). When the 168 SNPs in the inferred inversion were removed, this three-part structure disappeared and the second principal component was no longer significant (p = 0.09; Figure 2F).

383

384 SNP Selection and Genetic Diversity

Of the full 9,376 high-quality SNP set, 3,962 SNPs comprised 1,434 groups in LD with $R^2 \ge$ 385 0.8; all but one SNP was removed from each group to create a set of 6,848 "independent" 386 SNPs. This screening differs from the earlier LDna analysis, which sought to identify large 387 clusters of LD; by contrast, this approach intends simply to remove bias to population 388 structure measurements from including any sets of SNPs in strong LD. The majority of these 389 groups (N = 839) comprised small numbers of SNPs in the same contig. The largest of these 390 groups by far included 77 SNPs from the inferred inversion; no other groups contained >13 391 SNPs. We note that this analysis did not identify all 168 SNPs in the inferred inversion as 392 being in the same group, since some of those SNPs are in LD with others at $R^2 < 0.8$; 393

- consequently, the set of 6,848 independent SNPs includes 54 SNPs in the inferred inversion.
- 396 After removing SNPs in strong LD, we had a final working panel of 6,848 high-quality,
- independent SNPs. BayPass identified 537 of these SNPs as potentially under selection at
- ³⁹⁸ FDR ≤0.1 across all site-by-year samples; these SNPs were removed to construct a panel of
- 399 6,311 putatively neutral SNPs for selected downstream analyses.

400

Allelic richness ranged from 1.669-1.711 (Table 1); it was significantly lower in Seadrift Lagoon than in all other sites in all years ($p \le 0.005$; Table 1). Private allelic richness was low, ranging from 0.00438-0.00771; seven pairwise comparisons were significant, all comparing Seadrift Lagoon in 2011 or 2016 with non-Seadrift Lagoon sites (p < 0.05 for all comparisons). Lower allelic richness in Seadrift Lagoon reflects a lower number of polymorphic SNPs there than in all other sites (Table 1). All samples showed a significant heterozygote excess.

408

409 Population Structure and Migration

410 F_{ST} was significant between most sample pairs when using the 6,848 independent SNP set 411 (pairwise F_{ST} excluding temporal comparisons: 0.00058-0.027; SI Table S1). By contrast, when using the 6,311 putatively neutral SNPs nearly all significant comparisons were 412 413 between Seadrift Lagoon samples and all other sites (pairwise F_{ST} including Seadrift: 0.0081-0.016; pairwise F_{ST} excluding Seadrift: -0.0021-0.0044; SI Table S2). There was no evidence 414 for differentiation within any temporal comparison across years with the putatively neutral 415 SNP set (pairwise F_{ST}: -0.0020-0.0014; SI Table S2). Principal components analysis 416 reinforced these patterns, with the first component separating Seadrift Lagoon from all other 417 sites with both the independent and neutral SNP sets (6,848 independent SNPs: loading = 418 2.75%, p < 0.0001; 6,311 neutral SNPs: loading = 2.07%, p < 0.0001; Figure 3A,B). The 419 second component was significant only with the independent SNP set (loading = 1.53%, p < 420 0.0001), and spread non-Seadrift Lagoon sites along a rough north-south axis (Figure 3A). 421 With the neutral SNP set, this pattern collapsed (p > 0.05), with near-complete overlap among 422 all non-Seadrift Lagoon samples (Figure 3B). While Seadrift Lagoon had significantly lower 423 allelic richness than all other sites, there were no significant differences among the remaining 424

425 open sites (Figure 3C).

426

To test a realistic migration scenario, we estimated relative migration using only the most 427 recent sample from each site. Estimates of relative migration between sites (using 6,311 428 neutral SNPs only) demonstrated similar and symmetrical migration among all sites except 429 Seadrift Lagoon (Figure 3D). Consistent with Seadrift Lagoon's distinctiveness, we found 430 evidence for reduced migration both into and out of this site relative to the rest of our study 431 sites (Figure 3D). The approach we used sets the maximum observed migration to 1 and 432 433 scales the rest of the migration estimates accordingly. Between all five open sites, we observed values of 88-100% of maximum observed migration, while estimated migration 434 between Seadrift Lagoon and all other sites ranged from 68-78% of the maximum. We note, 435 however, that it is impossible to fully differentiate between ongoing low-level migration and 436 437 recent divergence (with no ongoing migration) given the recent history of green crabs in the northeast Pacific. 438

439

A test of IBD along the five "open" sites (excluding Seadrift Lagoon) recapitulated the north-440 441 south pattern observed for these sites in the PCA, but only when putatively selected SNPs were included. The 6,848 independent SNP set showed a significant pattern of IBD (R² = 442 0.38; p = 0.0002), which collapsed completely with the 6,311 neutral SNP set ($R^2 = -0.02$; p =443 0.4) or when removing only the 54 SNPs in the inferred inversion ($R^2 = 0.00$; p = 0.3; Figure 444 4A). However, we note that overall differentiation for all of these SNP sets was very low, with 445 a maximum pairwise F_{ST} of 0.0066. IBD was much stronger in the 144 outlier SNPs 446 (frequency outliers across the five open sites), and stronger still in the 54 "independent" SNPs 447 in the inferred inversion. The 144 frequency outliers had significant IBD with a maximum 448 pairwise F_{ST} of 0.16 (R^2 = 0.38; p = 0.002), while the SNPs in the inferred inversion showed a 449 strong IBD pattern ($R^2 = 0.77$; p < 0.0001) with a maximum pairwise F_{ST} of 0.24 between the 450 most distant two sites (Figure 4B). 451

452

453 Selection in the Northeast Pacific

All tests for selection were run with only the most recent temporal sample from the five open
sites, using the 6,662 SNPs of the 6,848 independent SNP set that were variable in these five

456 samples. Using the BayPass core model, 144 unlinked SNPs were frequency outliers at FDR 457 ≤0.05 among these five samples. The BayPass covariate test identified 26 SNPs related to 458 latitude, 26 SNPs associated with January SST, and six SNPs associated with July SST (SI 459 Figure S1). Seventeen of these SNPs were associated with two or more traits, for a total of 40 460 unique environmentally associated SNPs. Some associations were quite strong, with seven 461 SNPs associated with latitude and/or winter temperature at BF ≥ 25 dB. No SNPs were 462 strongly associated with July SST (SI Figure S1).

463

464 The Redundancy Analysis (RDA) showed that latitude fell almost perfectly along the first RDA axis, while July SST fell between the first and second axes (SI Figure S2). In this test, 465 association with latitude also represents association with January SST, which was not 466 included as the two measures were strongly correlated. The full RDA was significant (F = 467 1.10, p = 0.001), as were both resulting RDA axes (RDA1: F = 1.12, p = 0.001; RDA2: F = 468 1.09, p = 0.003). Variance Inflation Factors were less than 2 for both axes, indicating no 469 potential confounding from multicollinearity in the environmental variables. In total, 23 SNPs 470 were identified as outliers on one of the two RDA axes: of these, 15 were most strongly 471 472 associated with latitude while 8 were most strongly associated with July SST (SI Figure S2). 473

We took as our most likely candidate markers for selection those associated with latitude or 474 475 temperature in both the BayPass and RDA analyses. In total, 13 SNPs overlapped between 476 the 40 identified with BayPass and the 23 identified with RDA. Of these, two were excluded because of rarity (maximum per-population MAF \leq 0.3). Ten of the remaining 11 candidate 477 SNPs were in the inferred inversion; these were retained in the independent SNP set because 478 479 they were not in strong enough LD with each other to have been removed (threshold for strong LD: $R^2 \ge 0.8$). For visualization purposes, we generated "extended genotypes" for the 480 inferred inversion by classing individuals based on group membership in a PCA of all 168 481 SNPs (Figure 2C). As noted earlier, individuals belonging to the left-most group were classed 482 as homozygotes for the minor allele, those in the middle group as heterozygotes, and those in 483 the right-most group as homozygotes for the major allele. For the open sites (minus Seadrift 484 Lagoon), both the inferred inversion and the independent candidate SNP had MAFs that were 485 significantly correlated with latitude (Figure 5). Interestingly, for the inferred inversion, MAF in 486

Seadrift Lagoon fell outside the predicted line, instead having a MAF closer to that of British
Columbia and predicted to belong to a more northern and/or colder site (Figure 5A). By
contrast, Seadrift MAF fell along the line predicted by MAF in the open sites for the one
candidate SNP outside this inferred inversion (Figure 5B).

491

The ten candidate SNPs in the inferred inversion were in strong LD with a number of SNPs 492 removed from the unlinked data set, for a total of 97 SNPs in 39 different contigs (SI Table 493 S3). Because we ran selection analyses on a set of SNPs that had been pruned to remove 494 those in strong LD, any of these 97 SNPs (or other linked variation we did not retain after 495 SNP QC) could be driving the observed pattern of selection. Of the 97 candidate SNPs in the 496 inferred inversion, 18.6% (18/97) were predicted to change the amino acid sequence of the 497 resulting protein compared to 12.7% (1,192/9,374) in the full 9,376 SNP set (before linkage 498 499 filtering). The 18 predicted non-synonymous SNPs in the inferred inversion were in contigs annotated as: hypoxia inducible factor 1 alpha (2 SNPs), NAD-dependent protein deacylase, 500 fibrillin-2-like isoform X4, ubiquinol-cytochrome-c reductase complex assembly factor 1, SMB 501 domain-containing protein (3 SNPs), protein lingerer-like, prosaposin-like, cartilage oligomeric 502 503 matrix protein, ATP-dependent RNA helicase DDX56, one uncharacterized protein, and four 504 unannotated contigs (5 SNPs). The one candidate SNPs not in the inferred inversion was 505 predicted to be a synonymous substitution in protein SGT1 homolog (SI Table S4).

506

507 Discussion

508

The expansion of *C. maenas* along the northeast Pacific coast is a canonical example of the 509 "genetic paradox of invasions". The population has been demonstrably successful, rapidly 510 expanding along >1,500 km of coastline despite deriving from a single genetically 511 depauperate source. Here, we have shown that variation in a specific region of the genome – 512 an inferred chromosomal inversion previously associated with cold tolerance in the species – 513 appears to be under strong latitudinal selection in this system. Population genomics shows 514 that this putative selection is occurring against a backdrop of high oceanographic connectivity. 515 Our sampling comprises five discrete bays connected by high larval gene flow, and a single 516 oceanographically isolated population that has diverged genetically in <20 generations, 517

demonstrating the importance of larval connectivity in mediating population dynamics in range 518 expansion. Comparison with older data shows that genetic structure and diversity have 519 remained stable across at least one generation, suggesting large population sizes and 520 consistent recruitment pools over time. We propose that this high connectivity, a hallmark of 521 the species, may have promoted the initial evolution of this inferred inversion as a balanced 522 polymorphism in Europe and may be critical to its persistence and spread in introduced 523 populations. In turn, the variation protected by this inversion may play a key role in C. 524 maenas' success across wide environmental gradients in its introduced range despite 525 526 significant reductions in overall genetic diversity.

527

528 Chromosomal Inversions and Adaptation

The importance of chromosomal architecture, particularly chromosomal inversions, has been 529 increasingly recognized in selection with gene flow in natural systems (Tigano & Friesen, 530 2016). Inversion polymorphisms can be extensive, and can maintain extended genotypes at 531 hundreds to thousands of genes by suppressing recombination in heterozygotes (Kirkpatrick, 532 2010). While inversion is not the only mechanism by which recombination can be reduced, it 533 534 is generally believed to be most effective in maintaining large blocks of co-adapted genes 535 over time (Lamichhaney & Andersson, 2019). This recombination suppression, in turn, permits suites of gene variants to evolve and be inherited together, capturing complex multi-536 gene interactions in a single "supergene" (Thompson & Jiggins, 2014). 537

538

Inversions have been directly associated with important differences in ecotype across small 539 spatial scales in interbreeding populations, suggesting that this type of chromosomal 540 architecture can promote highly localized selection (Westram et al., 2018; Huang et al. 2020). 541 In some systems, including monkeyflowers and Atlantic cod, important differences in life 542 history have been linked to just one or two inversions (Twyford & Friedman 2015; Kirubakaran 543 et al. 2016). Pioneering work on Drosophila identified a number of inversions showing clinal 544 associations with latitude and temperature; such relationships have been shown to develop 545 rapidly in introduced populations (Balanyà et al. 2006), and to predictably "cycle" in frequency 546 with changing seasonal temperatures within a population (Kapun et al. 2016). Together, this 547 growing body of work suggests that inversions can act as targets of spatial balancing 548

selection in systems where the scale of gene flow exceeds the scale of environmental
heterogeneity, providing an effective mechanism by which such species can respond to their
environments on very fast time scales (Sanford & Kelly, 2011; Tigano & Friesen, 2016).

552

We have previously proposed that a chromosomal inversion or another genomic region of 553 reduced recombination is likely under selection to temperature in C. maenas (Tepolt & 554 Palumbi, 2020). Many of the same SNPs identified in the inferred inversion in this study were 555 independently found to be part of the putative inversion in that prior global study, indicating 556 that the same likely inversion is associated with cold tolerance globally and with latitudinal 557 divergence along the northeast Pacific. In the earlier study, candidate SNPs were variable in 558 the European native range and showed similar evidence of strong linkage disequilibrium and 559 reduced recombination there (Tepolt & Palumbi, 2020). This demonstrates that rapid selection 560 in the northeast Pacific is acting primarily on standing variation that arose in the original 561 source population, ruling out post-invasion dynamics as a driver for this linkage disequilibrium 562 (Slatkin, 2008). 563

564

565 Population Structure and Temporal Dynamics

In the northeast Pacific, C. maenas has lost significant overall genetic diversity compared to 566 its East Coast source or the species' native range in Europe (Tepolt & Palumbi, 2015). 567 However, diversity is largely consistent across its northeast Pacific range, with no losses at 568 569 the range edges relative to the San Francisco source (Figure 3C). While diversity loss in expanding range edges has been widely noted and is a common expectation (Vucetich, 570 Waite, & Nunney, 1997; Eckert et al., 2008), it may be mediated by large population sizes and 571 high gene flow (Excoffier, Foll, & Petit, 2009). Green crabs spread rapidly but episodically up 572 the coast from the initial point of introduction in San Francisco Bay, with the largest expansion 573 in conjunction with the strong 1997-1998 ENSO event (Behrens Yamada et al., 2005). Further 574 research has shown that strong crab cohorts have corresponded with warmer waters and 575 enhanced northward nearshore currents (Behrens Yamada, Peterson, & Kosro, 2015). 576 Modeling has shown that larval dispersal trajectories likely vary considerably both within and 577 between years depending on hydrography, with larvae potentially traveling both north and 578 south (Brasseale, Grason, McDonald, Adams, & MacCready, 2019). Together with this prior 579

work, our data suggest that periodic transport events are sufficient to maintain consistent
 genetic diversity and structure across time and space in these open bays despite the variable
 nature of recruitment in the system.

583

All of our data point to ongoing high gene flow across the range of the recent and rapid C. 584 *maenas* expansion in the northeast Pacific, with one exception (Figure 3). This exception is 585 Seadrift Lagoon, which is small, isolated from the adjacent Bolinas Lagoon, and now 586 oceanographically separated from the larger coastal circulation (Ritter, 1970). Green crabs 587 were first reported in Seadrift Lagoon in 1993 (Tepolt et al., 2009), and by 2011, our first year 588 of sampling, they had lost significant genetic diversity relative to the rest of our study sites 589 (Tepolt & Palumbi, 2015; Figure 3C). Our current sampling does not include Bolinas Lagoon, 590 the larger lagoon to which Seadrift Lagoon was historically connected but to which it is now 591 592 linked only by culverts with managed water flow. However, a prior study of the temporal dynamics of C. maenas using microsatellites found no evidence for diversity loss in Bolinas 593 Lagoon relative to any other sites, including San Francisco Bay and Bodega Bay (Tepolt et 594 al., 2009). Data from experimental removal work in Seadrift Lagoon suggest that population 595 596 dynamics in the lagoon are highly localized (Grosholz et al., 2021). Given the observed 597 openness of the other bays, we would expect structure and diversity to homogenize quickly if Seadrift Lagoon were receiving substantial larval inputs from surrounding, higher-diversity 598 599 populations.

600

Genetic structure and diversity appear to be stable over time at least for the 5-6 years covered by our sampling, with no significant changes in F_{ST} or allelic richness across years within a site. The lifespan of *C. maenas* is no more than 4-6 years (Behrens Yamada et al., 2005), and we did not sample the largest and oldest individuals, so the 2011 and 2016 samples represent non-overlapping generations. Structure and diversity were stable in both sites we sampled across generations, including one of the well-mixed open sites (Barkley Sound), and the putatively isolated Seadrift Lagoon.

608

609 High Gene Flow and Rapid Selection

610 Against a background of high gene flow and negligible neutral genetic structure among most

sites along the northeast Pacific, we observed a north-south gradient in the system driven by 611 an inferred inversion polymorphism (Figure 3A, 4). While it is very difficult to disentangle 612 selection from allele surfing at range expansion (Excoffier et al., 2009; Lotterhos & Whitlock, 613 2014), and we cannot say conclusively that allele surfing does not play a role in this IBD 614 pattern, several lines of evidence suggest that we are detecting a genuine signal of selection. 615 Green crabs along the northeastern Pacific coast comprise large populations with high 616 dispersal and gene flow, traits that limit the potential for successful allele surfing (Excoffier et 617 al., 2009; Goodsman, Cooke, Coltman, & Lewis, 2014). These traits are reflected in similar 618 Tevels of genetic diversity across all of the populations in the more highly connected "open" 619 bays (Figure 3C). In addition, while *C. maenas* has spread primarily northward from its site of 620 first introduction, the sole area of southern spread with an established population (Elkhorn 621 Slough) shows MAF consistent with increases of "southern" alleles. This is contrary to the 622 expectations of allele surfing, in which a species expanding along multiple range edges is 623 expected to demonstrate different "favored" alleles in each direction by chance (Demastes, 624 Hafner, Hafner, Light, & Spradling, 2019). 625

626

627 Finally, many of the same SNPs found in the inferred inversion driving latitudinal divergence were previously identified as belonging to a putative supergene strongly associated with 628 thermal physiology in a dataset spanning six native- and invasive-range C. maenas 629 630 populations (Tepolt & Palumbi, 2020). While winter SST and latitude cannot be disentangled in our current dataset, this previous study provided stronger evidence for temperature in 631 driving selection. Together with the minimal neutral structure across the open sites, we 632 suggest that the inferred inversion in our study is very likely maintained as a balanced 633 polymorphism under strong selection to temperature. 634

635

Recent research using fine-scale sampling covering multiple years, life stages, and sampling sites has shown that targets of selection can vary across all of these scales in high-dispersal systems, contributing to patterns that may appear chaotic with less thorough sampling (Thia et al., 2021). While "chaotic genetic patchiness" is often a hallmark of such systems (Eldon et al., 2016), we did not observe that in our sampling (Figure 3A,B; 5). This may be due in part to the domination of the selective signal in our system by a single large genomic region with

what is likely a high selection coefficient, in concert with our sampling of adults. If selection at
this region is acting primarily on the dispersive larval stage, our sampling will reflect the
aftermath of this selection rather than the initial pool of recruits (Sanford & Kelly, 2011).
Similar patterns of balanced polymorphism have been shown in two classic examples of
selection on large-effect alleles in early life stages in barnacles and mussels (Koehn et al.,
1980; Schmidt & Rand, 2001).

648

For SNPs in the inferred inversion, MAF at Seadrift Lagoon did not follow the predicted 649 650 relationship based on the open sites and was instead characteristic of higher latitudes (Figure 5A). While speculative, we suggest that Seadrift Lagoon's isolation means that crabs at this 651 site are responding to the environment on an extremely local scale as opposed to those in 652 other bays, whose larvae may travel hundreds of kilometers through coastal currents 653 654 (Behrens Yamada et al., 2005; Brasseale et al., 2019). Seadrift Lagoon is shallow and small, 655 and may experience more extreme (and especially colder) temperatures than nearby open bays. Finer-scale temperature data from within Seadrift Lagoon, rather than larger-scale 656 satellite-derived SST data, would help to test this hypothesis. 657

658

659 While prior work uncovered a robust link between this inferred inversion and physiological cold tolerance, its ability to identify rapid selection after invasion was limited by a complex 660 661 invasion history and differences in genetic background across the six studied populations on 662 three coastlines. Here, we demonstrate that this inferred inversion recapitulates predicted allele frequency correlation with temperature in an otherwise homogenous, highly 663 bottlenecked introduced population over a period of 10-20 generations. This study provides 664 evidence for very rapid adaptive change in an introduced species with extremely limited 665 genetic diversity, and proposes this adaptation was facilitated by variation at a single 666 inversion polymorphism that evolved and is likely maintained as a balanced polymorphism in 667 the native range. 668

669

670 Conclusions

671 We have long known that diversity is important to population resilience in the face of changing
672 conditions (Reed & Frankham, 2003). The genetic paradox of invasions is that we do

occasionally find incredibly successful, non-clonal populations that have passed through 673 severe bottlenecks, dramatically decreasing their genetic diversity relative to their sources 674 (Kohn, Murphy, Ostrander, & Wayne, 2006). Perhaps we can partially resolve this paradox by 675 considering that diversity at specific parts of the genome (rather than genome-wide diversity) 676 may play a critical role in resilience (Estoup et al., 2016). Simulations have shown that 677 expanding populations can adapt via a few variants of large effect even in the face of low 678 overall diversity (Gilbert & Whitlock, 2017). High dispersal, which characterizes many marine 679 systems, may promote the evolution of a few alleles of large effect via genomic mechanisms 680 such as inversion polymorphisms (Tigano & Friesen, 2016). While balanced polymorphisms 681 at large-effect alleles may permit these populations to respond extremely quickly to their local 682 environments, they may also be a huge benefit to the survival and success of introduced 683 populations. 684

686 Introduced marine species often exhibit an extensive dispersal ability, resulting from close association with human-built marine infrastructure or a high capacity for larval transport 687 (Carlton & Geller, 1993; Wilson, Dormontt, Prentis, Lowe, & Richardson, 2009). For the latter, 688 689 high dispersal and gene flow may have a twofold effect wherein the same traits that allow a 690 species to reach and spread in a new range may also promote the evolution of genomic 691 mechanisms (i.e., balanced polymorphisms) that facilitate rapid adaptation to a range of 692 environmental conditions (Tigano & Friesen, 2016). This is similar to the idea that periodic disturbance promotes the evolution of traits that enhance invasiveness and increase the 693 likelihood of success in novel environments (Lee & Gelembiuk, 2008; Ketola et al., 2013). We 694 propose that an analogous process may be at work in highly dispersive marine invaders. 695 Such species may be able to evolve and maintain balanced polymorphisms across broad 696 environmental gradients in their native ranges, giving them the substrate for rapid adaptive 697 change as they expand in new environments. 698

699

685

700 Acknowledgments

701

We thank S. Yamada, J. Gonzalez, R. Jeppeson, I. McGaw, and E. Clelland for their
assistance in obtaining genetic samples. We also thank the National Science Foundation

(OCE-RAPID #1514893 to EDG, CD and GM), Smithsonian Institution (Hunterdon Fund to
 GMR), and The Penzance Endowed Fund for Assistant Scientists (to CKT) for their support of
 this project.

707

708 References

- Balanyà, J., Oller, J. M., Huey, R. B., Gilchrist, G. W., & Serra, L. (2006). Global genetic
 change tracks global climate warming in Drosophila subobscura. *Science*, 313, 1773–
 1775. https://doi.org/10.1126/science.1131002
- Behrens Yamada, S., & Gillespie, G. (2008). Will the European green crab (*Carcinus maenas*) persist in the Pacific Northwest? *ICES Journal of Marine Science*, 65, 725–729.
 https://doi.org/10.1093/Icesjms/Fsm191
- Behrens Yamada, S., & Kosro, P. (2010). Linking ocean conditions to year class strength of
 the invasive European green crab, *Carcinus maenas*. *Biological Invasions*, 12, 1791–
- 717 1804. https://doi.org/10.1007/s10530-009-9589-y
- Behrens Yamada, S., Peterson, W., & Kosro, P. (2015). Biological and physical ocean
 indicators predict the success of an invasive crab, *Carcinus maenas*, in the northern
 California Current. *Marine Ecology Progress Series*, 537, 175–189.
- 721 https://doi.org/10.3354/meps11431
- Behrens Yamada, S., Dumbauld, B. R., Kalin, A., Hunt, C. E., Figlar-Barnes, R., Randall, A.
 (2005). Growth and persistence of a recent invader *Carcinus maenas* in estuaries of the
 northeastern Pacific. *Biological Invasions*, 7, 309–321. https://doi.org/10.1007/s10530004-0877-2
- Benjamini, Y., & Hochberg, Y. (1995). Controlling the false discovery rate: a practical and
 powerful approach to multiple testing. *Journal of the Royal Statistical Society B*, 57, 289–300.
- Blackburn, T. (2008). Using aliens to explore how our planet works. *Proceedings of the National Academy of Sciences U.S.A.* 105, 9–10.
- 731 https://doi.org/10.1073/pnas.0711228105
- Brasseale, E., Grason, E., McDonald, P. S., Adams, J., & MacCready, P. (2019). Larval

- transport modeling support for identifying population sources of European green crab in
 the Salish Sea. *Estuaries and Coasts*, 42, 1586–1599. https://doi.org/10.1007/s12237019-00586-2
- Bridle, J., & Vines, T. (2007). Limits to evolution at range margins: when and why does
 adaptation fail? *Trends in Ecology and Evolution*, 22, 140–147.
 https://doi.org/10.1016/j.tree.2006.11.002
- Bors, E., Herrera, S., Morris, J., & Shank, T. (2019). Population genomics of rapidly invading
 lionfish in the Caribbean reveals signals of range expansion in the absence of spatial
 population structure. *Ecology and Evolution*, 9, 3306–3320.
- 742 https://doi.org/10.1002/ece3.4952
- Carlton, J., & Cohen, A. (2003). Episodic global dispersal in shallow water marine organisms:
 the case history of the European shore crabs *Carcinus maenas* and *C. aestuarii. Journal of Biogeography*, 30, 1809–1820. https://doi.org/10.1111/j.1365-2699.2003.00962.x
- Carlton, J., & Geller, J. (1993). Ecological roulette: the global transport of nonindigenous
 marine organisms. *Science*, 261, 78–82.
- Chuang, A., & Peterson, C. (2016). Expanding population edges: theories, traits, and tradeoffs. *Global Change Biology*, 22, 494–512. https://doi.org/10.1111/gcb.13107
- Csardi, G., & Nepusz, T. (2006). The igraph software package for complex network research,
 InterJournal, Complex Systems, 1695. http://igraph.org
- Danecek, P., Auton, A., Abecasis, G., Albers, C. A., Banks, E., Depristo, M. A., ... 1000
 Genome Project Analysis Group. (2011). The variant call format and VCFtools. *Bioinformatics*, 27, 2156–2158.
- Dawirs, R. (1985). Temperature and larval development of *Carcinus maenas* (Decapoda) in
 the laboratory; predictions of larval dynamics in the sea. *Marine Ecology-Progress Series*, 24, 297–302. https://doi.org/10.3354/meps024297
- Demastes, J., Hafner, D., Hafner, M., Light, J., & Spradling, T. (2019). Loss of genetic
 diversity, recovery and allele surfing in a colonizing parasite, *Geomydoecus aurei*. *Molecular Ecology*, 28, 703–720. https://doi.org/10.1093/oxfordjournals.jhered.a111627

- DePristo, M., Banks, E., Poplin, R., Garimella, K. V., Maguire, J. R., Hartl, C. ... Daly, M. J.
 (2011) A framework for variation discovery and genotyping using next-generation DNA sequencing data. *Nature Genetics*, 43, 491–498.
- de Rivera, C., Hitchcock, N. G., Teck, S. J., Steves, B. P., Hines, A. H., Ruiz, G. M. (2007).
 Larval development rate predicts range expansion of an introduced crab. *Marine Biology*, 150, 1275–1288. https://doi.org/10.1007/s00227-006-0451-9
- Dowle, M., & Srinivasan, A. (2019). data.table: Extension of `data.frame`. R package version
 1.12.8. https://CRAN.R-project.org/package=data.table
- Eckert, C., Samis, K., & Lougheed, S. (2008). Genetic variation across species' geographical
 ranges: the central-marginal hypothesis and beyond. *Molecular Ecology*, 17, 1170–1188.
 https://doi.org/10.1111/j.1365-294X.2007.03659.x
- Eldon, B., Riquet, F., Yearsley, J., Jollivet, D., & Broquet, T. (2016). Current hypotheses to
 explain genetic chaos under the sea. *Current Zoology*, 62, 551–566.
- 774 https://doi.org/10.1093/cz/zow094
- Ellingson, R., & Krug, P. (2016). Reduced genetic diversity and increased reproductive
 isolation follow population-level loss of larval dispersal in a marine gastropod. *Evolution*,
 70, 18–37. https://doi.org/10.1111/evo
- Estoup, A., Ravigné, V., Hufbauer, R., Vitalis, R., Gautier, M., & Facon, B. (2016). Is there a
 genetic paradox of biological invasion? *Annual Review of Ecology, Evolution, and Systematics*, 47, 51–72. https://doi.org/10.1146/annurev-ecolsys-121415-032116
- Excoffier, L., Foll, M., & Petit, R. (2009). Genetic consequences of range expansions. *Annual Review of Ecology, Evolution, and Systematics*, 40, 481–501.
- 783 https://doi.org/0.1146/annurev.ecolsys.39.110707.173414
- Excoffier, L., & Lischer, H. E. L. (2010). Arlequin suite ver 3.5: A new series of programs to
 perform population genetics analyses under Linux and Windows. *Molecular Ecology Resources*, *10*, 564–567.
- Excoffier, L., & Ray, N. (2008). Surfing during population expansions promotes genetic
 revolutions and structuration. *Trends in Ecology and Evolution*, 23, 347–351.

789	https://doi.org/10.1016/j.tree.2008.04.004
790 791 792	Forester, B., Lasky, J., Wagner, H., Urban, D. (2018). Comparing methods for detecting multilocus adaptation with multivariate genotype–environment associations. <i>Molecular Ecology</i> , <i>27</i> , 2215–2233.
793 794 795	Galindo, H., Olson, D., & Palumbi, S. (2006). Seascape genetics: A coupled oceanographic- genetic model predicts population structure of Caribbean corals. <i>Current Biology</i> , 16, 1622–1626. https://doi.org/10.1016/j.cub.2006.06.052
796 797	Gautier, M. (2015). Genome-wide scan for adaptive divergence and association with population-specific covariates. <i>Genetics, 201</i> , 1555–1579.
798 799 800	Gilbert, K., & Whitlock, M. (2017). The genetics of adaptation to discrete heterogeneous environments: frequent mutation or large-effect alleles can allow range expansion. <i>Journal of Evolutionary Biology</i> , 30, 591–602. https://doi.org/10.1111/jeb.13029
801 802 803 804	Goodsman, D., Cooke, B., Coltman, D., & Lewis, M. (2014). The genetic signature of rapid range expansions: how dispersal, growth and invasion speed impact heterozygosity and allele surfing. <i>Theoretical Population Biology</i> , 98, 1–10. https://doi.org/10.1016/j.tpb.2014.08.005
805 806 807 808	 Grosholz, E., Ashton, G., Bradley, M., Brown, C., Ceballos-Osuna, L., Chang, A., Tepolt, C. (2021). Stage-specific overcompensation, the Hydra effect, and the failure to eradicate an invasive predator. <i>Proceedings of the National Academy of Science U.S.A.</i>, 118, e2003955118. https://doi.org/10.1073/pnas.2003955118
809 810 811	Hampton, S., & Griffiths, C. (2007). Why <i>Carcinus maenas</i> cannot get a grip on South Africa's wave-exposed coastline. <i>African Journal of Marine Science</i> , 29, 123–126. https://doi.org/10.2989/AJMS.2007.29.1.11.76
812 813 814	 Hart, A., Ginzburg, S., Xu, M., Fisher, C. R., Rahmatpour, N., Mitton, J. B., Wegrzyn, J. L. (2020). EnTAP: Bringing faster and smarter functional annotation to non-model eukaryotic transcriptomes. <i>Molecular Ecology Resources, 20</i>, 591–604.
815 816	Hedgecock, D. (1986). Is gene flow from pelagic larval dispersal important in the adaptation and evolution of marine invertebrates? <i>Bulletin of Marine Science</i> , 39, 550–564.

817	Hida	algo, F., Barón, P., & Orensanz, J. (2005). A prediction come true: the green crab invades
818		the Patagonian coast. Biolical Invasions, 7, 547–552. https://doi.org/10.1007/s10530-
819		004-5452-3

Huang, K., Andrew, R. L., Owens, G. L., Ostevik, K. L., & Rieseberg, L. H. (2020). Multiple
 chromosomal inversions contribute to adaptive divergence of a dune sunflower ecotype.
 Molecular Ecology, 29, 2535–2549. https://doi.org/10.1111/mec.15428

- Kapun, M., Fabian, D., Goudet, J., & Flatt, T. (2016). Genomic evidence for adaptive
 inversion clines in *Drosophila melanogaster*. *Molecular Biology & Evolution*, 33, 1317–
 1336. https://doi.org/10.1093/molbev/msw016
- Kawecki, T., Ebert, D. (2004). Conceptual issues in local adaptation. *Ecology Letters*, 7,
 1225–1241. https://doi.org/10.1111/j.1461-0248.2004.00684.x

Kemppainen, P., Knight, C. G., Sarma, D. K., Hlaing, T., Prakash, A., Maung Maung, Y. N., ...
 Walton, C. (2015). Linkage disequilibrium network analysis (LDna) gives a global view of
 chromosomal inversions, local adaptation and geographic structure. *Molecular Ecology Resources*, 15, 1031–1045. https://doi.org/10.1111/1755-0998.12369

Ketola, T., Mikonranta, L., Zhang, J., Saarinen, K., Örmälä, A. M., Friman, V., ... Laakso, J.

(2013). Fluctuating temperature leads to evolution of thermal generalism and

preadaptation to novel environments. *Evolution*, 67, 2936–2944.

835 https://doi.org/10.1111/evo.12148

Kilkenny, F., & Galloway, L. (2012). Adaptive divergence at the margin of an invaded range. *Evolution*, 67, 722–731. https://doi.org/10.5061/dryad.6b2t6

Kinlan, B., & Gaines, S. (2003). Propagule dispersal in marine and terrestrial environments: a
 community perspective. *Ecology*, 84, 2007–2020. https://doi.org/10.1890/01-0622

- Kirkpatrick, M. (2010). How and why chromosome inversions evolve. *PloS Biology*, 8,
 e1000501. https://doi.org/10.1371/journal.pbio.1000501
- Kirubakaran, T. G., Grove, H., Kent, M. P., Sandve, S. R., Baranski, M., Nome, T., ...
 Andersen, Ø. (2016). Two adjacent inversions maintain genomic differentiation between
 migratory and stationary ecotypes of Atlantic cod. *Molecular Ecology*, 25, 2130–2143.

845 https://doi.org/10.1111/mec.13592

Koehn, R., Newell, R., & Immermann, F. (1980). Maintenance of an aminopeptidase allele
 frequency cline by natural selection. *Proceedings of the National Academy of Sciences U.S.A.*, 77, 5385–5389. https://doi.org/10.1073/pnas.77.9.5385

Kohn, M., Murphy, W., Ostrander, E., & Wayne, R. (2006). Genomics and conservation

genetics. *Trends in Ecology and Evolution*, 21, 629–637.

851 https://doi.org/10.1016/j.tree.2006.08.001

- Lamichhaney, S., & Andersson, L. (2019). A comparison of the association between large
 haplotype blocks under selection and the presence/absence of inversions. *Ecology and Evolution*, 9, 4888–4896. https://doi.org/10.1002/ece3.5094
- Langmead, B., & Salzberg, S. (2013). Fast gapped-read alignment with Bowtie 2. *Nature Methods, 9,* 357–359.
- Lee, C., & Gelembiuk, G. (2008). Evolutionary origins of invasive populations. *Evolutionary Applications*, 1, 427–448. https://doi.org/10.1111/j.1752-4571.2008.00039.x
- Lee, C., Kiergaard, M., Gelembiuk, G., Eads, B., & Posavi, M. (2011). Pumping ions: Rapid
 parallel evolution of ionic regulation following habitat invasions. *Evolution*, 65, 2229–
 2244. https://doi.org/10.1111/j.1558-5646.2011.01308.x
- Llaurens, V., Whibley, A., & Joron, M. (2017). Genetic architecture and balancing selection:
 the life and death of differentiated variants. *Molecular Ecology*, 26, 2430–2448.
 https://doi.org/10.1111/mec.14051
- Lotterhos, K. E., & Whitlock, M. C. (2014). Evaluation of demographic history and neutral
 parameterization on the performance of F_{ST} outlier tests. *Molecular Ecology*, 23, 2178–
 2192. https://doi.org/10.1111/mec.12725
- Martin, M. (2011). Cutadapt removes adapter sequences from high-throughput sequencing
 reads. EMBnet Journal, 17, 10–12.
- McKenna, A., Hanna, M., Banks, E., Sivachenko, A., Cibulskis, K., Kernytsky, A., ... DePristo,
 M. A. (2010) The Genome Analysis Toolkit: a MapReduce framework for analyzing nextgeneration DNA sequencing data. Genome Resources, 20, 1297–1303.

	873 874	n, X. J., Butler, G., Storms, R., & Tsang, A. (2005). OrfPredictor: predicting protein-coding regions in EST-derived sequences. <i>Nucleic Acids Research</i> , 33, W677–W680.						
	875	Munday, P., Warner, R., Monro, K., Pandolfi, J., & Marshall, D. (2013). Predicting evolutionary						
	876 877	https://doi.org/10.1111/ele.12185						
878 879 880		 Oksanen, J., Blanchet, F. G., Friendly, M., Kindt, R., Legendre, P., McGlinn, D., Wagner, H. <i>vegan: Community Ecology Package</i> (R package version 2.4-3. https://CRAN.R-project.or). R package version 2.4-3. https://CRAN.R-project.org. 						
	881 882 883	 Palumbi, S., & Pinsky, M. (2014). Marine dispersal, ecology, and conservation. <i>In</i> M. Bertness, J. Bruno, B. Silliman, & J. Stachowicz (Eds.), <i>Marine Community Ecology and Conservation</i> (1st ed., pp. 57–84). Sunderland, MA, USA: Sinauer Associates. 						
	884 885	Patro, R., Duggal, G., Love, M. I., Irizarry, R. A., & Kingsford, C. (2017). Salmon provides fast and bias-aware quantification of transcript expression. <i>Nature Methods</i> , <i>14</i> , 417–419.						
	886 887 888	Pembleton, L. W., Cogan, N. O. I., & Forster, J. W. (2013). StAMPP: An R package for calculation of genetic differentiation and structure of mixed-ploidy level populations. <i>Molecular Ecology Resources</i> , 13, 946–952.						
	889 890 891	Pespeni, M., & Palumbi, S. (2013). Signals of selection in outlier loci in a widely dispersing species across an environmental mosaic. <i>Molecular Ecology</i> , 22, 3580–3597. https://doi.org/10.1111/mec.12337						
	892 893	Phillips, B., Brown, G., Webb, J., & Shine, R. (2006). Invasion and the evolution of speed in toads. <i>Nature</i> , 439, 803–803. https://doi.org/10.1038/439803a						
	894 895 896	 Price, A., Patterson, N. J., Plenge, R. M., Weinblatt, M. E., Shadick, N. A., & Reich, D. (2006). Principal components analysis corrects for stratification in genome-wide association studies. <i>Nature Genetics</i>, <i>38</i>, 904–909. 						
	897 898 899 900	Prunier, J., Dubut, V., Chikhi, L., & Blanchet, S. (2017). Contribution of spatial heterogeneity in effective population sizes to the variance in pairwise measures of genetic differentiation. <i>Methods in Ecology and Evolution</i> , 8, 1866–1877. https://doi.org/10.1111/2041-210X.12820						

901 902	R Core Team. R: A Language and Environment for Statistical Computing. Retrieved from https://www.r-project.org
903 904	Reed, D., & Frankham, R. (2003). Correlation between fitness and genetic diversity. <i>Conservation Biology</i> , 17, 230–237. https://doi.org/10.1046/j.1523-1739.2003.01236.x
905 906 907	Reynolds, R. W., Smith, T. M., Liu, C., Chelton, D. B., Casey, K. S., & Schlax, M. G. (2007). Daily high-resolution-blended analyses for sea surface temperature. <i>Journal of Climate</i> , <i>20</i> , 5473–5496.
908 909	Ritter, J. (1970). A summary of preliminary studies of sedimentation and hydrology in Bolinas Lagoon, Marin County, California. U.S. Geological Survey, Circular 627.
910 911 912	Rius, M., & Darling, J. (2014). How important is intraspecific genetic admixture to the success of colonising populations? <i>Trends in Ecology and Evolution</i> , 29, 233–242. https://doi.org/10.1016/j.tree.2014.02.003
913 914 915	Roman, J., & Darling, J. (2007). Paradox lost: genetic diversity and the success of aquatic invasions. <i>Trends in Ecology and Evolution</i> , 22, 454–464. https://doi.org/10.1016/j.tree.2007.07.002
916 917	Rousset, F. (2008). genepop'007: a complete re-implementation of the genepop software for Windows and Linux. Molecular Ecology Resources, 8, 103–106.
918 919	Sanford, E., & Kelly, M. (2011). Local adaptation in marine invertebrates. <i>Annual Review of Marine Science</i> , 3, 509–535. https://doi.org/10.1146/annurev-marine-120709-142756
920 921 922	Schmidt, P., Bertness, M., & Rand, D. (2000). Environmental heterogeneity and balancing selection in the acorn barnacle, <i>Semibalanus balanoides</i> . <i>Philosophical Transactions of the Royal Society B</i> , 267, 379–384. https://doi.org/10.1111/j.0014-3820.2001.tb00656.x
923 924 925	Schmidt, P. S., & Rand, D. M. (2001). Adaptive maintenance of genetic polymorphism in an intertidal barnacle: Habitat- and life-stage-specific survivorship of Mpi genotypes. <i>Evolution</i> , 55, 1336–1344. https://doi.org/10.1111/j.0014-3820.2001.tb00656.x
926 927 928	See, K. & Feist, B. (2010). Reconstructing the range expansion and subsequent invasion of introduced European green crab along the west coast of the United States. <i>Biological Invasions</i> , 12, 1305–1318. https://doi.org/10.1007/s10530-009-9548-7

929 930 931	Sexton, J., Strauss, S., & Rice, K. (2011). Gene flow increases fitness at the warm edge of a species' range. <i>Proceedings of the National Academy of Sciences U.S.A.</i> , 108, 11704–11709. https://doi.org/10.1073/pnas.1100404108
932 933 934	Simms, D., Cizdziel, P., & Chomczynski, P. (1993). TRIzol: a new reagent for optimal single- step isolation of RNA. <i>Focus</i> , 99–102. https://doi.org/http://dx.doi.org/10.1016/0003- 2670(61)80041-X
935 936 937	Slatkin, M. (2008). Linkage disequilibrium–understanding the evolutionary past and mapping the medical future. <i>Nature Reviews Genetics</i> , 9, 477–485. https://doi.org/10.1038/nrg2361
938 939 940	Sotka, E. (2012). Natural selection, larval dispersal, and the geography of phenotype in the sea. <i>Integrative and Comparative Biology</i> , 52, 538–545. https://doi.org/10.1093/icb/ics084
941 942	Sundqvist, L., Keenan, K., Zackrisson, M., Prodöhl, P., & Kleinhans, D. (2016). Directional genetic differentiation and asymmetric migration. <i>Ecology & Evolution, 6</i> , 3461–3475.
943 944	Szpiech, Z. A., Jakobsson, M., & Rosenberg, N. A. (2008). ADZE: a rarefaction approach for counting alleles private to combinations of populations. <i>Bioinformatics</i> , <i>24</i> , 2498–2504.
945 946 947 948	 Szücs, M., Vahsen, M. L., Melbourne, B. A., Hoover, C., Weiss-Lehman, C., & Hufbauer, R. A. (2017). Rapid adaptive evolution in novel environments acts as an architect of population range expansion. <i>Proceedings of the National Academy of Sciences U.S.A.</i>, 114, 13501–13506. https://doi.org/10.1073/pnas.1712934114
949 950 951	 Takahashi, Y., Suyama, Y., Matsuki, Y., Funayama, R., Nakayama, K., & Kawata, M. (2016). Lack of genetic variation prevents adaptation at the geographic range margin in a damselfly. <i>Molecular Ecology</i>, 25, 4450–4460. https://doi.org/10.1111/mec.13782
952 953 954 955 956	 Tepolt, C., Darling, J. A., Bagley, M. J., Geller, J. B., Blum, M. J., & Grosholz, E. D. (2009). European green crabs (<i>Carcinus maenas</i>) in the Northeastern Pacific: genetic evidence for high population connectivity and current-mediated expansion from a single introduced source population. <i>Diversity and Distributions</i>, 15, 997–1009. https://doi.org/10.1111/j.1472-4642.2009.00605.x
	929 930 931 932 933 934 935 936 937 938 939 940 941 942 943 944 945 944 945 946 947 948 945 946 947 948 949 945 946 947 948 949 950 951 952 953 954 955

Tepolt, C., & Palumbi, S. (2015). Transcriptome sequencing reveals both neutral and adaptive
 genome dynamics in a marine invader. *Molecular Ecology*, 24, 4145–4158.
 https://doi.org/10.1111/mec.13294

Tepolt, C., & Palumbi, S. (2020). Rapid adaptation to temperature via a potential genomic
 island of divergence in the invasive green crab, *Carcinus maenas*. *Frontiers in Ecology and Evolution*, 8, 580701. https://doi.org/10.3389/fevo.2020.580701

Tepolt, C., & Somero, G. (2014). Master of all trades: thermal acclimation and adaptation of
 cardiac function in a broadly distributed marine invasive species, the European green
 crab, *Carcinus maenas*. *Journal of Experimental Biology*, 217, 1129–1138.
 https://doi.org/10.1242/jeb.093849

Thia, J., McGuigan, K., Liggins, L., Figueira, W. F., Bird, C. E., Mather, A., ... Riginos, C.
 (2021). Genetic and phenotypic variation exhibit both predictable and stochastic patterns
 across an intertidal fish metapopulation. *Molecular Ecology*, In press.

- 970 https://doi.org/s0740-5472(96)90021-5
- Thompson, M. J., & Jiggins, C. D. (2014). Supergenes and their role in evolution. *Heredity*,
 113, 1–8. https://doi.org/10.1038/hdy.2014.20

Tigano, A., & Friesen, V. (2016). Genomics of local adaptation with gene flow. *Molecular Ecology*, 25, 2144–2164. https://doi.org/10.1111/mec.13606

⁹⁷⁵ Twyford, A. D., & Friedman, J. (2015). Adaptive divergence in the monkey flower *Mimulus*

guttatus is maintained by a chromosomal inversion. *Evolution*, 69, 1476–1486.

977 https://doi.org/10.5061/dryad.5h032.

Véliz, D., Duchesne, P., Bourget, E., & Bernatchez, L. (2006). Stable genetic polymorphism in
 heterogeneous environments: Balance between asymmetrical dispersal and selection in
 the acorn barnacle. *Journal of Evolutionary Biology*, 19, 589–599.

981 https://doi.org/10.1111/j.1420-9101.2005.01000.x

Vucetich, J., Waite, T., & Nunney, L. (1997). Fluctuating population size and the ratio of
 effective to census population size. *Evolution*, 51, 2017–2021.

984 Weir, B. S., & Cockerham, C. C. (1984). Estimating *F*-statistics for the analysis of population

structure. Evolution, 38, 1358–1370. 985 Welch, W. (1968). Changes in abundance of the green crab, Carcinus maenas (L.), in 986 relation to recent temperature changes. Fishery Bulletin, 67, 337-345. 987 Westram, A., Rafajlović, M., Chaube, P., Faria, R., Larsson, T., Panova, M., ... Butlin, R. 988 (2018). Clines on the seashore: the genomic architecture underlying rapid divergence in 989 the face of gene flow. Evolution Letters, 2-4, 297-309. https://doi.org/10.1002/evl3.74 990 White, C., Selkoe, K. A., Watson, J., Siegel, D. A., Zacherl, D. C., & Toonen, R. J. (2010). 991 992 Ocean currents help explain population genetic structure. Proceedings of the Royal Society B, 277, 1685–1694. https://doi.org/10.1098/rspb.2009.2214 993 White, T., Perkins, S., Heckel, G., & Searle, J. (2013). Adaptive evolution during an ongoing 994 range expansion: the invasive bank vole (Myodes glareolus) in Ireland. Molecular 995 Ecology, 22, 2971–2985. https://doi.org/10.1111/mec.12343 996 Wickham, H. (2016). ggplot2: Elegant Graphics for Data Analysis. Retrieved from 997 998 http://ggplot2.org Wilson, J., Dormontt, E., Prentis, P., Lowe, A., & Richardson, D. (2009). Something in the way 999 you move: dispersal pathways affect invasion success. Trends in Ecology and Evolution, 1000 24, 136–144. https://doi.org/10.1016/j.tree.2008.10.007 1001 Xuereb, A., Benestan, L., Normandeau, É., Daigle, R. M., Curtis, J. M. R., Bernatchez, L., & 1002 Fortin, M. J. (2018). Asymmetric oceanographic processes mediate connectivity and 1003 population genetic structure, as revealed by RADseq, in a highly dispersive marine 1004 invertebrate (Parastichopus californicus). Molecular Ecology, 27, 2347–2364. 1005 1006 https://doi.org/10.1111/mec.14589 1007 **Data Accessibility** 1008 Raw transcriptome reads from new sequencing for this project have been deposited in 1009 GenBank's Sequence Read Archive (SRA) under BioProject ID PRJNA690934 and 1010 BioSample IDs SAMN17267686-SAMN17267781. The cleaned transcriptome, high-quality 1011 individual SNP genotypes, and custom scripts used in this bioinformatics pipeline from in this 1012

- 1013 paper will be archived in Dryad upon acceptance. Raw sequence reads from 2011,
- reanalyzed in this project, are available in the SRA under BioProject ID PRJNA283611 and
- 1015 BioSample IDs SAMN03653390-SAMN03653413.
- 1016

1017 Author Contributions

All authors designed the research. CT performed research, analyzed data, and wrote the paper. All authors edited multiple drafts.

Tables

Table 1: Sampling information and summary statistics for each site x year sample. Adult = 1+ year old animal; YOY = Young of the Year or <1 year old animal. N_{poly} = number of polymorphic SNPs; Ar = allelic richness; pAr = private allelic richness; H_0 = observed heterozygosity; H_E = expected heterozygosity; F_{IS} = Wright's inbreeding coefficient. All samples showed a significant heterozygote excess (p < 0.05).

Code	Site	Coordinates	Year	Age	Ν	N _{poly}	Ar	pAr	Ho	HE	Fıs
ES_15*	Elkhorn Slough, CA	36.820, -121.745	2015 A	Adult	12	5,546	1.708	0.00697	0.2612	0.2510	-0.0409
SF_15*	San Francisco Bay, CA	37.997, -122.370	2015 A	Adult	12	5,549	1.702	0.00621	0.2510	0.2491	-0.0078
SF_16	San Francisco Bay, CA	37.997, -122.370	2016	YOY	12	5,560	1.705	0.00666	0.2593	0.2501	-0.0365
SL_11	Seadrift Lagoon, CA	37.907, -122.662	2011 A	Adult	12	5,233	1.669	0.00494	0.2461	0.2414	-0.0196
SL_15*	Seadrift Lagoon, CA	37.907, -122.662	2015	YOY	12	5,234	1.675	0.00584	0.2609	0.2424	-0.0761
SL_16	Seadrift Lagoon, CA	37.907, -122.662	2016	YOY	12	5,264	1.677	0.00434	0.2498	0.2442	-0.0228
BB_15*	Bodega Bay, CA	38.324, -123.054	2015 A	Adult	12	5,538	1.706	0.00645	0.2681	0.2511	-0.0674
OR_16*	Tillamook Bay, OR	45.542, -123.904	2016	YOY	12	5,548	1.711	0.00762	0.2618	0.2539	-0.0311
BC_11	Barkley Sound, BC	49.026, -125.346	2011 A	Adult	12	5,562	1.711	0.00771	0.2600	0.2520	-0.0316
BC_16*	Barkley Sound, BC	49.026, -125.346	2016 A	Adult	12	5,517	1.705	0.00676	0.2630	0.2512	-0.0471

* indicates samples used in tests of selection and relative migration.

Figures

Figure 1: Map of sampling sites; year(s) of sampling given in italics under site names. From north to south: Barkley Sound, BC (2011, 2016); Tillamook Bay, OR (2016); Bodega Bay, CA (2015); Seadrift Lagoon, CA (2011, 2015, 2016); San Francisco Bay, CA (2015, 2016); and Elkhorn Slough, CA (2015). Asterisk indicates the location where the species was initially introduced (first detected in 1989).



Figure 2: Linkage disequilibrium analysis of the full 9,376 SNP set, showing evidence for an inferred inversion. A-B: Clustering in LDna network analysis showing nested single and compound outlier LD clusters. In B, the inferred inversion (compound cluster in A) is indicated by an arrow and highlighted in red. C: PCA of 168 SNPs in inferred inversion. D: F_{IS} of 168 SNPs in inferred inversion, grouped by PC1 position in C. E: PCA of the full 9,376 SNP set. F: PCA of the full 9,376 SNP set excluding the 168 SNPs in the inferred inversion. For C-F, four individuals were excluded because they were missing high-quality genotypes at >20% of SNPs in the inferred inversion.



Figure 3: Genetic structure, diversity, and migration across all sites, showing high gene flow among all sites except Seadrift Lagoon. A: PCA of the independent 6,848 SNPs. B: PCA of the putatively neutral 6,311 SNPs. C: Allelic richness by population and collection year across the 6,848 independent SNPs, plotted against distance from the initial point of introduction in San Francisco Bay, CA. Points from the same site have been jittered horizontally for clarity. Vertical bars indicate standard error. Starred samples have significantly lower allelic richness than non-starred samples. D: Relative migration between sites, calculated with the Nm method, across the 6,662 independent SNPs in the five "open" sites. The highest observed rate is set to 1, and all other rates are scaled to that maximum; estimated migration both into and out of Seadrift was lower than migration between all other sites.



Figure 4: Comparison of isolation-by-distance (IBD) patterns across all open populations (excluding Seadrift Lagoon), showing IBD driven by SNPs in the inferred inversion. A: For all 6,848 independent SNPs (red); 6,794 independent SNPs excluding the large outlier LD cluster (gray); and 6,311 putatively neutral independent SNPs (black). B: For all 144 frequency outlier SNPs across the open populations (aqua); and all 54 SNPs from the inferred inversion in the independent SNP set (blue). The relationships in A are shown in B with smaller points; note the difference in Y-axis magnitude between the two panels.



Figure 5: Correlation between Minor Allele Frequency (MAF) and latitude or July SST at candidate genomic regions, showing putative selection to temperature or latitude across the five "open" sites at environmental outlier SNPs. Asterisks indicate the location where the species was initially introduced (first detected in 1989). In both cases, Seadrift Lagoon (gray point) is shown for comparison but was not used in the regression lines or equations. A) The inferred inversion; samples were classed into overall "inversion genotypes" based on their group membership in Figure 2C. B) Outlier SNP in the protein SGT homolog transcript.

